

RESEARCH ARTICLE

Measuring the Short-term Success of Grassland Restoration: The Use of Habitat Affinity Indices in Ecological Restoration

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Abstract

Agricultural intensification threatens grasslands worldwide and the restoration of grasslands from arable lands can at least partially counter this threat. We studied grassland restoration by following early successional changes of arthropod assemblages (spiders, Araneae; true bugs, Heteroptera; orthopterans, Orthoptera; and ground beetles, Carabidae) on 1- and 2-year-old restorations using arable lands and native grasslands as two ends of the succession timescale. To examine the changes in species composition among the habitat types, we used habitat affinity indices based on fidelity and/or specificity of the species. We found that the number of species did not differ between habitat types, while species composition changed markedly with time. Species richness was thus not adequate to

detect favorable changes after grassland restoration. Habitat affinity indices, on the other hand, were useful to detect compositional changes caused by the increasing numbers of species characteristic of target grasslands as early as the second year after restoration. Habitat affinity indices are easy-to-use, easy-to-interpret measures of restoration success; therefore, we recommend their use as measures complementary to species richness and simple similarity. Our results show that sowing low-diversity seed mixture followed by mowing and grazing can be particularly successful in grassland restoration in time periods as short as 2 years.

Key words: arthropods, fidelity, large-scale restoration, low-diversity seed mixture, specificity.

Introduction

Agricultural intensification threatens grasslands worldwide and is one of the main reasons for grassland restoration in Europe (Bakker & Berendse 1999; Muller 2002; Pywell et al. 2002; Walker et al. 2004). Intensive agriculture often causes habitat loss and degradation, decreases the number of specialist species and increases the number of generalists, which results in biotic homogenization (McKinney & Lockwood 1999). To address the loss of biodiversity in agro-ecosystems, many EU countries have introduced agri-environmental schemes (Muller 2002; Kleijn & Sutherland 2003; Walker et al. 2004). Furthermore, because the small number and restricted area of remaining grasslands apparently limit our ability to preserve grassland biodiversity (Shepherd & Debinski 2005; Billeter et al. 2008), the restoration of grasslands has come into focus in many countries (Cramer et al. 2008). The abandonment

of fields currently ongoing in some countries provides good opportunities for the conversion of arable lands to grasslands to create new grassland patches, enlarge existing patches or to enhance connectivity between patches (Hobbs & Cramer 2007).

Conservation actions treated as experiments and followed up by monitoring help us to gain a better understanding of the ecological processes induced by restoration (Lindenmayer et al. 2008). The most commonly used measures to follow habitat changes after restoration are species richness and diversity indices of selected taxa (Perner & Malt 2003; Piper et al. 2007). However, species richness can be misleading because it does not always correlate with conservation objectives or ecological function (Borrwall & Ebenman 2008). A strongly disturbed, early successional habitat, for example, can host a large number of pioneer and disturbance-tolerant species, while successional processes often involve a decrease in total species richness together with an increase in the richness of the taxa characteristic to the target habitats (e.g. Paquin 2008; Török et al. 2008). In such cases, the changes in species composition cannot be detected with overall species richness and other measures need to be used.

Local- or small-scale restorations are over-represented in the literature (Henry et al. 2008; Wagner et al. 2008). Existing

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large-scale restorations are rarely monitored (Petersen et al. 2003; Wagner et al. 2008). Furthermore, multi-taxon monitoring is also uncommon (Ruiz-Jaen & Aide 2005; Woodcock et al. 2008). Conservation actions affect a wide range of animal taxa (e.g. Perner & Malt 2003; Purtauf et al. 2004; Kardol et al. 2005; Piper et al. 2007; Woodcock et al. 2008), and the joint monitoring of plants and animals is essential to measure the general success of restoration (George & Zack 2001; Legg & Nagy 2006; Miller & Hobbs 2007; Lengyel et al. 2008). Vegetation monitoring is suggested to perform better on long-term assessments, while invertebrates can be useful to detect short-term effects of restoration because they respond quickly to microclimatic changes (Kremen et al. 1993; Mattoni et al. 2000; Perner & Malt 2003).

The aim of our study was to follow short-term changes in arthropod assemblages after landscape-scale grassland restoration and to examine the effectiveness of habitat affinity indices in detecting these changes. The success of converting arable land to grassland is often limited by seed dispersal or high soil fertility (Pywell et al. 2002; Walker et al. 2004; Kardol et al. 2008). Therefore, different methods are used in grassland restoration to counteract those effects. Here we applied the most frequently used practice by sowing seed mixtures of target grass species after soil preparation and by mowing and grazing the fields from the first year after restoration. However, in contrast to most previous restorations that used seed mixtures of 10–40 species, we used low-diversity seed mixtures (two or three species), which provided more room for natural colonization processes. To measure the short-term effectiveness of grassland restoration, we chose several groups of vegetation- and ground-dwelling arthropods (spiders, true bugs, orthopterans, and ground beetles). In addition to changes in species richness, we also assessed changes in species composition, by using nonmetric multidimensional scaling ordination and recently developed habitat affinity indices (Tóthmérész & Magura 2005; Magura et al. 2006). We expected that all the used statistical methods (species richness, ordination, and habitat affinity indices) would converge to show that the arthropod assemblages of the restored grasslands are moving toward those of the target native grasslands with succession. The modified habitat affinity indices and ordination were more effective than species richness in assessing the early successional changes of arthropod assemblages because they take into account the identity of the species and their affinities to certain habitats, including the habitats targeted by restoration.

Methods

Study Area and Sampling

The study was carried out in the Egyek-Pusztakócs marsh and grassland system (N 47°33', E 20°54'), a 50 km² unit of Hortobágy National Park, the oldest (1973) and largest (17,000 km²) national park in Hungary. The ongoing landscape-scale restoration project implements several different conservation actions (e.g. grassland restoration, grazing,

fire management) to protect and restore two Natura 2000 priority habitat types (Pannonic alkali steppes and marshes, Pannonic loess steppic grasslands). Grassland restoration by sowing low-diversity seed mixtures (two or three grass species for alkali and loess grasslands, respectively) has taken place on approximately 5 km² arable land in 2005–2007. We surveyed four different habitat types in 2007: (1) arable lands, which were either grain or alfalfa fields; (2) 1-year-old restored grasslands (fields sown in 2006), representing a pioneer successional stage; (3) 2-year-old restored grasslands (sown in 2005), representing a latter stage of succession; and (4) native grasslands, either alkali or loess steppic grasslands that were designated as the targets of the restoration.

One-year-old grasslands were dominated mostly by herbaceous, short-lived weedy plants (e.g. *Capsella bursa-pastoris*, *Matricaria inodora*), and some weedy grasses (e.g. *Bromus arvensis*, *Bromus mollis*) to a lesser extent. On 2-year-old grasslands, weeds were replaced by perennial graminoid species (e.g. *Festuca pratensis*, *Elymus hispidus*) and some species characteristic of target grasslands also appeared (e.g. *Achillea collina*, *Convolvulus arvensis*). The target, species-poor alkali grasslands (*Festucion pseudovinae*) are dominated by the grass *Festuca pseudovina*, while *Poa angustifolia*, *E. hispidus*, *Carex stenophylla* are subordinate graminoid species. Other typical herbaceous species include *Achillea setacea*, *A. collina*, and *Trifolium* species. The target loess steppic grasslands (*Festucion rupicolae*) are dominated by several grasses (e.g. *Festuca rupicola*, *F. valesiaca*, *P. angustifolia*, *Bromus inermis*), and are rich in herbaceous species including *Salvia nemorosa*, *Galium verum*, *Dianthus pottederae*, and *Euphorbia cyparissias* with some rare and protected plants, including *Phlomis tuberosa*, *Thalictrum minus*, and *Ornithogalum pyramidale* (Török et al. 2009, in press).

In total, 39 plots were studied (5 arable lands, 10 one-year-old, 11 two-year-old, and 13 native grasslands) between May and September in 2007. Plot area was between 0.04 and 0.36 km² (mean: 0.16 km²). The plots were at least 200 m away from each other. We surveyed vegetation- and ground-dwelling spiders (Araneae), true bugs (Heteroptera), orthopterans (Orthoptera), and ground beetles (Carabidae) using identical methods on each plot. We have selected these arthropod groups to include both herbivore and predator trophic levels and to include both vegetation- and ground-dwelling life forms. Ground-dwelling invertebrates were sampled by pitfall traps installed in two randomly selected locations within every plot in May. Traps were 0.5 L plastic cups with 100 mL ethylene-glycol (75%) as killing liquid, and were covered by fiberboard to keep out vertebrates, and to avoid contamination by litter or rain. Pitfall traps were placed at least 50 m from each other and from the edges of the plots. Vegetation-dwelling invertebrates were sampled by standardized sweepnetting consisting of 200 strokes in 150-m-long transects that started from the pitfall traps and progressed in randomly selected directions. Invertebrates were collected once every 3 weeks, on a total of six occasions during the vegetation period (May—September) to account for phenological changes in arthropod assemblages.

Specimens were identified to species in the lab. For all analyses, we pooled samples from the six occasions. Changes in vegetation were monitored by botanists, who collected data on plant species richness and diversity, plant cover and phytomass. Details on the changes of vegetation are discussed elsewhere (Deák et al. 2008; Török et al. 2009, in press).

Data Analyses

We studied whether arthropod assemblages have changed with secondary succession after restoration. Allegro and Sciaky (2002) proposed the forest affinity index to assess the relative quality of a given habitat compared to another habitat using species frequencies. Based on their work, Magura et al. (2006) developed three new habitat affinity indices based on habitat specificity and/or fidelity because they found that an affinity index using only relative frequencies is biased toward assemblages containing few, highly abundant “less-specialized” species. Habitat affinity indices based on fidelity and/or specificity emphasize commonness and rarity (Tóthmérész & Magura 2005). Because newly developed indices proved to be useful in comparing different types of forests (Magura et al. 2006), we have adapted them to grasslands. Here we used affinity indices based on specificity (HAS), fidelity (HAF), and both specificity and fidelity combined (HAFS).

We defined the species’ affinity values (A_i) in a four-step procedure. First, we classified the plots by hierarchical cluster analysis (Bray–Curtis dissimilarity index and Ward fusion method). The cluster analysis classified the plots in well-defined habitat groups (Fig. 1). Second, we assigned values ranging from -1 to $+1$ to each habitat type and to groups of habitat types (Fig. 1). Third, we performed an indicator value analysis (IndVal) as proposed by Dufrene and Legendre (1997), which assigned species to habitats that they are characteristic of based on the observed patterns of occurrence and abundance. Finally, we determined the affinity values of the species based on IndVal. Affinity values for species could thus take the following seven values:

- +1: species characteristic of target grasslands;
- +0.75: species related to 2-year-old restored grasslands and target grasslands;
- +0.5: species characteristic of 2-year-old restored grasslands;
- 0: species indifferent to habitat type (habitat generalists);
- 0.5: species related to 1-year-old restored grasslands;
- 0.75: species characteristic of 1-year-old restored grasslands and arable lands;
- 1: species characteristic of arable lands.

The exact values of affinity do not affect the results as long as the ordinal nature of affinities (most favorable species receiving highest score) is maintained. We have repeated all analyses presented in the paper by using different arbitrary values of the ordinal affinity variable and the results were qualitatively similar to those presented here. We present results by using 0 for those species which have no particular habitat preference and positive values for species of conservation

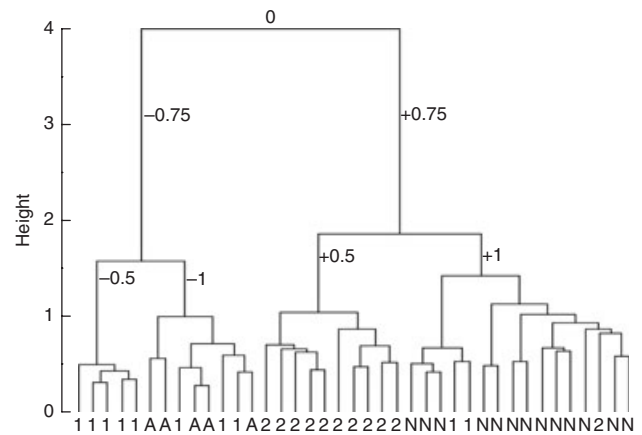


Figure 1. Dendrogram obtained in a hierarchical cluster analysis of surveyed plots with habitat affinity values assigned to each habitat type and groups of habitat types. Labels: 1, 1-year-old restored grasslands; 2, 2-year-old restored grasslands; A, arable lands; N, native grasslands. Affinity values given are shown on the dendrogram: -1 , species characteristic to arable lands; -0.75 , species characteristic to 1-year-old restored grasslands and arable lands; -0.5 , species characteristic to 1-year-old restored grasslands; 0, species indifferent to habitat type (habitat generalists); $+0.5$, species characteristic to 2-year-old restored grasslands; $+0.75$, species related to 2-year-old restored grasslands and target grasslands; $+1$, species characteristic to target grasslands.

importance and species identified by IndVal as characteristic of the target habitats. This logic follows the original suggestions of Allegro and Sciaky (2002).

When all species were given an affinity value, we calculated the affinity indices of the sampling plots. The habitat affinity index based on fidelity (HAF) was calculated as follows:

$$HAFr = \sum_{i=1}^S (I_{ir} \cdot \pi_i \cdot A_i) \quad (1)$$

where S is the total number of species; I_{ir} is the indicator function, which is either 0 or 1 depending on whether the species is present in plot r or not; π_i is the relative frequency of plots where species i is present in a given habitat type; and A_i is the value of habitat affinity of species i .

The habitat affinity index based on specificity (HAS) was calculated as follows:

$$HASr = \sum_{i=1}^S (I_{ir} \cdot e_i \cdot A_i) \quad (2)$$

where e_i is the specificity of the species i . Specificity is defined as the ratio of the average number of individuals in the given habitat type compared to the average number of individuals across all habitats.

Finally, the habitat affinity index based on both fidelity and specificity (HAFS) is calculated as follows:

$$HAFSr = \sum_{i=1}^S (I_{ir} \cdot \sqrt{\pi_i \cdot e_i} \cdot A_i) \quad (3)$$

with notations the same as above. The values of the indices are higher if more species characteristic of the target habitat are present in the habitat.

Analyses of variance (ANOVA) were performed to detect the differences in the mean value of the habitat affinity indices among habitat types (arable lands, 1-year-old, 2-year-old, native grasslands). If there were significant differences among habitat types, Tukey's HSD (Honestly Significant Difference) were used to compare means. Data were checked to meet the assumptions of ANOVA. We used two-sided tests and $\alpha = 0.05$ significance levels in statistical tests. Nonmetric multidimensional scaling ordination (with Bray–Curtis dissimilarity index) based on abundance data of the collected species was used to display changes in arthropod species composition after grassland restoration (Legendre & Legendre 1998). All statistical analyses were performed using R version 2.5.1 (R Development Core Team 2007).

Results

A total of 341 species (with 17,199 individuals) were collected, identified and given affinity (A_i) values. Most species belonged to true bugs (96 species, 7,424 individuals) and vegetation-dwelling spiders (82 species, 1,498 individuals), followed by ground beetles and ground-dwelling spiders (67 and 66 species, 6,305 and 1,492 individuals, respectively), and orthopterans were the least speciose (30 species, 480 individuals) taxon.

The average number of species was roughly similar in the three grassland habitats (Table 1; Fig. 2). However, native grasslands had higher habitat affinity values than all restored sites indicating the high number of species characteristic to the target habitats. Two-year-old grasslands had higher values than the group of 1-year-old grasslands and arable lands, which did not differ from each other (Table 1; Fig. 3). Both arable lands and 1-year-old grasslands were dominated by disturbance-tolerant species. Although the HAS index (which takes into account the specificity of the species) appeared most sensitive to differences among habitat types, the differences were robust regardless of which index was used (Fig. 3). One-year-old grasslands had proportionally more species receiving higher A_i values than arable lands and 2-year-old grasslands had proportionally more of these species than did 1-year-old grasslands.

The nonmetric multidimensional scaling ordination showed that species composition changed with time. Arthropod assemblages of restored grasslands became progressively more similar to those of native grasslands (Fig. 4), indicating the increasing abundance of species characteristic of grasslands. The disturbed, pioneer environment in the first year favored mostly disturbance-tolerant arthropods (e.g. *Dictyna arundinacea* [Dictynidae, Araneae]; *Pardosa agrestis* [Lycosidae, Araneae]; *Trigonotylus ruficornis* [Miridae, Heteroptera]; *Chorthippus biguttulus* [Acrididae, Orthoptera]; *Pseudoophonus rufipes* [Carabidae, Coleoptera]). The changes in species composition between the first and the second year

Table 1. Results of ANOVA analyses testing differences in the number of species and the three habitat affinity indices based on fidelity (HAF), specificity (HAS), and both (HAFS) among arable lands ($n = 5$ plots), 1-year-old ($n = 10$), 2-year-old ($n = 11$) restored grasslands and native grasslands ($n = 13$).

Variable	<i>f</i> -values (<i>df</i> = 3,38)	Tukey HSD
Species richness	1.92 n.s.	Arable=1 year=2 years=ative
HAF (fidelity)	148.32***	(Arable=1 year)<2 years<ative
HAS (specificity)	104.01***	(Arable=1 year)<2 years<ative
HAFS (both)	142.60***	(Arable=1 year)<2 years<ative

n.s. = not significant;

Species richness did not differ by habitat types, while habitat affinity values were significantly higher in 2-year-old grasslands than on arable lands or 1-year-old grasslands, and native grasslands had the highest habitat affinity value.

*** $p < 0.001$.

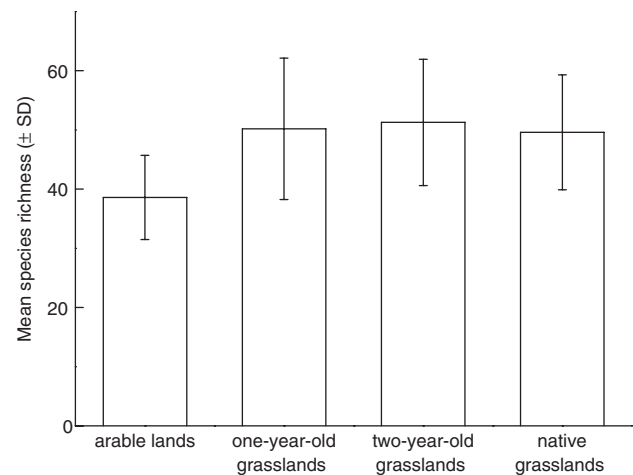


Figure 2. Mean (\pm SD) of species richness in the studied habitats. No significant differences were found between habitat types.

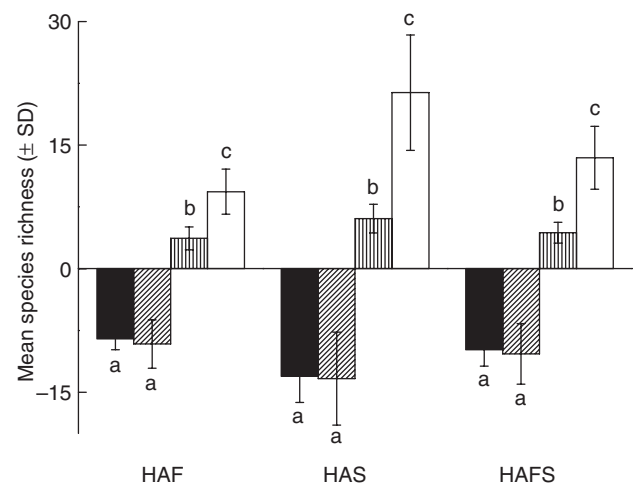


Figure 3. Mean (\pm SD) of habitat affinity indices based on fidelity (HAF), specificity (HAS), and both (HAFS). Legend: filled bars, arable lands; cross striped bars, 1-year-old restored grasslands; vertical striped bars, 2-year-old restored grasslands; open bars, native grasslands. Different letters indicate significant differences ($p < 0.05$).

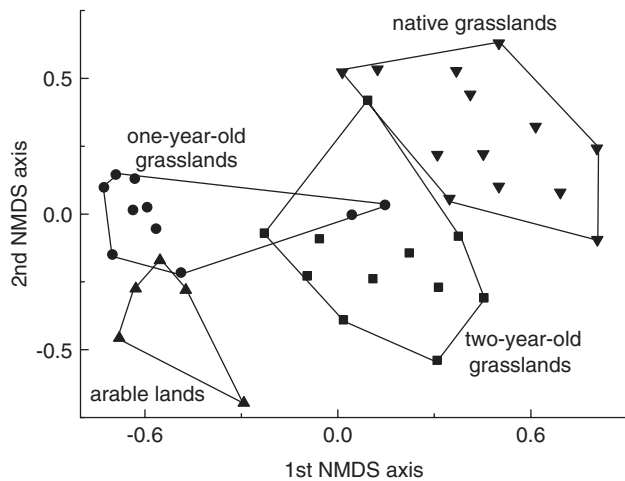


Figure 4. Changes in arthropod species composition after grassland restoration from arable lands (upward triangles) through 1-year-old (circles) and 2-year-old (squares) grasslands. Native grasslands are shown for reference (downward triangles).

after restoration were striking. The appearance of arthropods typical of grasslands (e.g. *Nesticus cellulans* [Nesticidae, Araneae]; *Titanoeca veteranica* [Titanocidae, Araneae]; *Chorosoma schillingi* [Rhopalidae, Heteroptera]; *Euchorthippus pulvinatus* [Acrididae, Orthoptera]; *Amara fulvipes* [Carabidae, Coleoptera]), caused a change in species composition as early as the second year after restoration. A detailed list of the species identified by the IndVal analysis as characteristic of a habitat type is given in the Appendix.

Discussion

The changes in arthropods detected here correspond well to changes in the vegetation of the sites following restoration. Similar to other grassland restoration projects (Van der Putten et al. 2000; Pywell et al. 2002; Camill et al. 2004), a vegetation composed almost entirely of weed plant species (e.g. *Capsella bursa-pastoris*, *Matricaria inodora*) appeared in the first year in our restored sites (Deák et al. 2008; Török et al. 2009, in press). Weeds might have facilitated germination and early development of the target grassland species (Callaway & Walker 1997; Pywell et al. 2002; Pueyo et al. 2009). Mowing favors the sown grasses that became superior competitors following the facilitative interaction (Callaway & Walker 1997). As a result, our sites had a strong grassland character as early as the second year after restoration (Deák et al. 2008; Török et al. 2009, in press).

The changes in vegetation were followed quickly by changes in the arthropod assemblages. The composition of arthropod assemblages of the restored sites was quickly approaching that of native grasslands. The changes in species composition between the first and the second year after restoration were striking and we initially expected these changes to occur later based on other restoration studies (Mortimer

et al. 2002; Purtauf et al. 2004). We believe that former alfalfa fields and native grasslands near to the restorations were the sources of the rapid arthropod recolonization (e.g. Shepherd & Debinski 2005). In our study, quick appearance of several species characteristic of the target grasslands in the 2-year-old restorations suggested that alfalfa fields, which are not plowed for 3–4 years, might provide refuges for grassland species and can thus serve as sources of colonization in restorations.

Our results are similar to those of other studies that used species richness integrated with some measure of species composition (e.g. diversity, evenness, similarity) to compare successional stages of grassland ecosystems (plants and butterflies: Steffan-Dewenter & Tscharntke 1997; springtails: Brand & Dunn 1998; plants, spiders, and beetles: Perner & Malt 2003; carabid beetles: Purtauf et al. 2004; Shepherd & Debinski 2005). We found that arthropod species richness did not change in the first 2 years following grassland restoration and did not differ significantly among successional stages. The increase of the values of the affinity indices with time showed that arthropod assemblages had changed due to the replacement of species indifferent to habitat type with species characteristic of target grasslands. Our results from using the indices corresponded with those of Magura et al. (2006) who examined ground beetle assemblages in Norway spruce (*Picea abies*) plantations. Magura et al. (2006) found that beech forests (natural ecosystem) had significantly higher affinity values than spruce plantations of different ages using the indices based on fidelity and/or specificity. Habitat affinity indices, although rarely used, are useful complementary estimates to the most common diversity measures. We encourage restoration ecologists to apply these indices in monitoring as they are easy to use and are easy to interpret.

Usually, more than a decade is necessary until the biodiversity of restored fields reaches that of the reference ecosystem (Brand & Dunn 1998; Purtauf et al. 2004). This process is particularly slow in the case of dry grasslands (Stadler et al. 2007) such as those targeted by our restoration. We believe that the rapid changes we observed may have occurred due to the proximity of propagule sources and/or the high availability of propagules. Our study suggests that the recovery of the restored fields can be accelerated by sowing foundation species only (Hutchings & Booth 1996a; Walker et al. 2004; Piper et al. 2007) or by applying different management actions (e.g. grazing, mowing, burning) after restoration (Hutchings & Booth 1996b). Although we agree that restoration is a long-term process and further monitoring is needed to follow subsequent changes, the quicker-than-expected positive changes in the arthropod assemblages observed here argue against views that restoration takes too much time to produce any results and that it is a waste of money to invest in ecological restoration projects (Aronson et al. 2006). Based on our study and several others (e.g. McCoy & Mushinsky 2002; Ruiz-Jaen & Aide 2005; Wilsey et al. 2005), we suggest that the use of multiple measurements (affinity indices, multivariate statistics, etc.) alongside with species richness is an effective way to assess the value of habitat restoration.

Implications for Practice

- Habitat affinity indices are good measures of restoration success. They take into account the identity of species, such that changes in species composition can be readily interpreted.
- Dense weed cover in the first year after restoration with low-diversity seed mixtures can provide a diverse habitat for arthropods. After mowing, rapid changes in the vegetation occur that produce changes in arthropod assemblages. The composition of restored assemblages approaches that of native grasslands as early as the second year after restoration.
- Proximity of semi-natural or native grasslands to restored fields can facilitate recolonization of arthropods.

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Appendix. List of significant ($p < 0.05$) character species of habitat types based on IndVal analysis. The affinity value (A_i) and the indicator value of the species, and the habitat types they are characteristic of are shown in the table. Indicator value is 100%, if the species occurs in every samples of a particular habitat type and it occurs only in that habitat type.

Species name	A_i value	Habitat type	IndVal (%)
Araneae (collected with pitfall traps)			
<i>Alopecosa pulverulenta</i>	1	Native grasslands	61.54
<i>Drassyllus praeficus</i>	1	Native grasslands	34.50
<i>Hogna radiata</i>	1	Native grasslands	30.77
<i>Micaria albobittata</i>	1	Native grasslands	23.08
<i>Oedothorax apicatus</i>	–0.5	One-year-old grasslands	44.76
<i>Pardosa agrestis</i>	0	Habitat indifferent	84.62
<i>Pardosa prativaga</i>	1	Native grasslands	40.67
<i>Tibellus maritimus</i>	0.5	Two-year-old grasslands	39.51
<i>Tibellus oblongus</i>	0.75	Two-year-old & native grasslands	46.58
<i>Trochosa robusta</i>	1	Native grasslands	38.46
<i>Zelotes hermani</i>	1	Native grasslands	23.08
<i>Zelotes latreillei</i>	1	Native grasslands	30.77
<i>Zelotes longipes</i>	1	Native grasslands	48.62
Araneae (collected with sweep-nets)			
<i>Enoplognatha mordax</i>	0.75	Two-year-old & native grasslands	39.09
<i>Erigone dentipalpis</i>	–1	Arable lands	62.51

Appendix. Continued

Species name	A_i value	Habitat type	IndVal (%)
<i>Hypsosinga heri</i>	1	Native grasslands	38.46
<i>Hypsosinga pygmaea</i>	-1	Arable lands	46.42
<i>Larinioides suspicax</i>	0.75	Two-year-old & native grasslands	36.36
<i>Nematogmus sanguinolentus</i>	-1	Arable lands	40.00
<i>Neoscona adianta</i>	0.5	Two-year-old grasslands	67.34
<i>Neottiura bimaculata</i>	0.5	Two-year-old grasslands	37.31
<i>Neriere radiata</i>	-1	Arable lands	33.49
<i>Pardosa palustris</i>	1	Native grasslands	23.08
<i>Tetragnatha extensa</i>	-1	Arable lands	50.42
<i>Xysticus kochi</i>	0.75	Two-year-old & native grasslands	56.34
<i>Xysticus striatipes</i>	1	Native grasslands	41.07
Heteroptera			
<i>Acetropis carinata</i>	0.5	Two-year-old grasslands	87.23
<i>Aelia acuminata</i>	0.75	Two-year-old & native grasslands	51.92
<i>Agramma articapillum</i>	1	Native grasslands	30.77
<i>Agramma confusum</i>	1	Native grasslands	76.92
<i>Camptobrochis punctulatus</i>	-0.5	One-year-old grasslands	52.91
<i>Capsodes gothicus</i>	-1	Arable lands	40.00
<i>Cymus glandicolor</i>	-0.5	One-year-old grasslands	58.96
<i>Emblethis verbasci</i>	-0.5	One-year-old grasslands	41.70
<i>Eurydema oleraceum</i>	-0.5	One-year-old grasslands	48.89
<i>Henestaris halophilus</i>	-0.5	One-year-old grasslands	67.26
<i>Heterogaster affinis</i>	-0.5	One-year-old grasslands	63.93
<i>Ischnodemus sabuleti</i>	1	Native grasslands	44.86
<i>Liocoris tripustulatus</i>	-0.75	Arable lands & 1-year-old grasslands	64.97
<i>Megalocera recticornis</i>	0.5	Two-year-old grasslands	31.47
<i>Nabis fesus</i>	-1	Arable lands	75.53
<i>Notostira erratica</i>	0.5	Two-year-old grasslands	64.61
<i>Nysius senecionis</i>	-0.5	One-year-old grasslands	30.00
<i>Orius minutus</i>	-0.5	One-year-old grasslands	38.87
<i>Orthotylus flavosparsus</i>	0.75	Two-year-old & native grasslands	56.02
<i>Piesma maculatum</i>	0.5	Two-year-old grasslands	60.00
<i>Polymerus cognatus</i>	-0.5	One-year-old grasslands	72.18
<i>Polymerus unifasciatus</i>	-0.5	One-year-old grasslands	45.64
<i>Rhopalus parumpunctatus</i>	-0.5	One-year-old grasslands	53.09
<i>Stenodema calcaratum</i>	0.75	Two-year-old & native grasslands	90.91
<i>Stenodema laevigatum</i>	-1	Arable lands	35.72
<i>Stictopleurus abutilon</i>	-0.5	One-year-old grasslands	64.66
<i>Stictopleurus punctatonervosus</i>	0.75	Two-year-old & native grasslands	36.41
<i>Syromastes rhombeus</i>	0.75	Two-year-old & native grasslands	36.36
<i>Xanthochilus quadratus</i>	1	Native grasslands	30.77
<i>Zicroma caerulea</i>	-0.5	One-year-old grasslands	58.63
Orthoptera			
<i>Chorthippus oschei</i>	0.5	Two-year-old grasslands	68.83
<i>Chorthippus parallelus</i>	0.5	Two-year-old grasslands	20.00
<i>Conocephalus discolor</i>	1	Native grasslands	48.83
<i>Decticus verrucivorus</i>	0.5	Two-year-old grasslands	45.83
<i>Euchorthippus declivus</i>	1	Native grasslands	78.70
<i>Omocestus haemorrhoidalis</i>	1	Native grasslands	30.77
<i>Omocestus rufipes</i>	1	Native grasslands	92.00
<i>Stenobothrus crassipes</i>	1	Native grasslands	46.15
<i>Tetrix subulata</i>	1	Native grasslands	30.77
<i>Mantis religiosa</i> (Ordo: Dictyoptera)	1	Native grasslands	30.77
Carabidae			
<i>Amara similata</i>	-0.5	One-year-old grasslands	49.14
<i>Anchomenus dorsalis</i>	-0.75	Arable lands & 1-year-old grasslands	40.00
<i>Anisodactylus signatus</i>	-0.75	Arable lands & 1-year-old grasslands	66.67
<i>Brachinus crepitans</i>	-0.5	One-year-old grasslands	71.55
<i>Brachinus explodens</i>	-0.75	Arable lands & 1-year-old grasslands	33.47
<i>Brachinus ganglbaueri advena</i>	-0.5	One-year-old grasslands	48.06
<i>Brachinus psophia</i>	-0.5	One-year-old grasslands	36.64

Appendix. Continued

<i>Species name</i>	A_i value	<i>Habitat type</i>	<i>IndVal (%)</i>
<i>Calosoma auropunctatum</i>	-0.5	One-year-old grasslands	30.00
<i>Cicindela germanica</i>	-1	Arable lands	49.96
<i>Dolichus halensis</i>	-0.75	Arable lands & 1-year-old grasslands	40.00
<i>Harpalus distinguendus</i>	0.75	Two-year-old & native grasslands	64.67
<i>Harpalus pygmaeus</i>	-0.75	Trable lands & 1-year-old grasslands	26.67
<i>Ophonus azureus</i>	-0.5	One-year-old grasslands	50.00
<i>Poecilus cupreus</i>	-0.75	Arable lands & 1-year-old grasslands	62.59
<i>Poecilus punctulatus</i>	-1	Arable lands	35.56
<i>Pseudoophonus calceatus</i>	-1	Arable lands	40.93
<i>Pseudoophonus rufipes</i>	-0.75	Arable lands & 1-year-old grasslands	95.55
<i>Pterostichus macer</i>	1	Native grasslands	53.95
<i>Pterostichus melanarius</i>	-0.75	Arable lands & 1-year-old grasslands	20.00
<i>Pterostichus ovoideus</i>	1	Native grasslands	28.32